



# **Changes in coastal environment and mass occurrence of jellyfish in Tokyo Bay**

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# Ecosystem in hyper eutrophicated bay

- High concentrations of nutrients (Inflow from river and drain, supply from bottom layer by mixing)
- High conc. N & P; Low conc. Si
- Microflagellates prevail over diatoms
- Production of the inedible algae enters the energy flow via detritus food chain

DOC → Bacteria → Heterotrophic nanoflagellates  
→ Ciliates (Microzooplankton)

Detritus Food Chain





# Problems accompanied with mass occurrence of jellyfish

- Decrease in fishing activities by clogging and bursting trawl nets

(Accumulation of dead jellyfish on bottom)

— *Aurelia aurita*, *Chrysaora melanaster*,  
*Nemopilema nomurai*

- Coastal power plants by blocking intakes for cooling water

— *Aurelia aurita*, Ctenophora

- Toxins

— Cubomedusae

# Mass occurrence of large jellyfish (*Nemopilema nomurai*) in Japan Sea



Photo in Awashima-Island

Mass occurrence of *N. nomurai* is observed in  
1920, 1958, 1995, 2002, 2003 & 2005



**Surface aggregation of *Aurelia aurita*  
in Tokyo Bay**

# Transition of Ecosystem

## Muscle Food Chain

Diatoms → Mesozooplankton → Fish

(Observed mainly in upwelling systems)

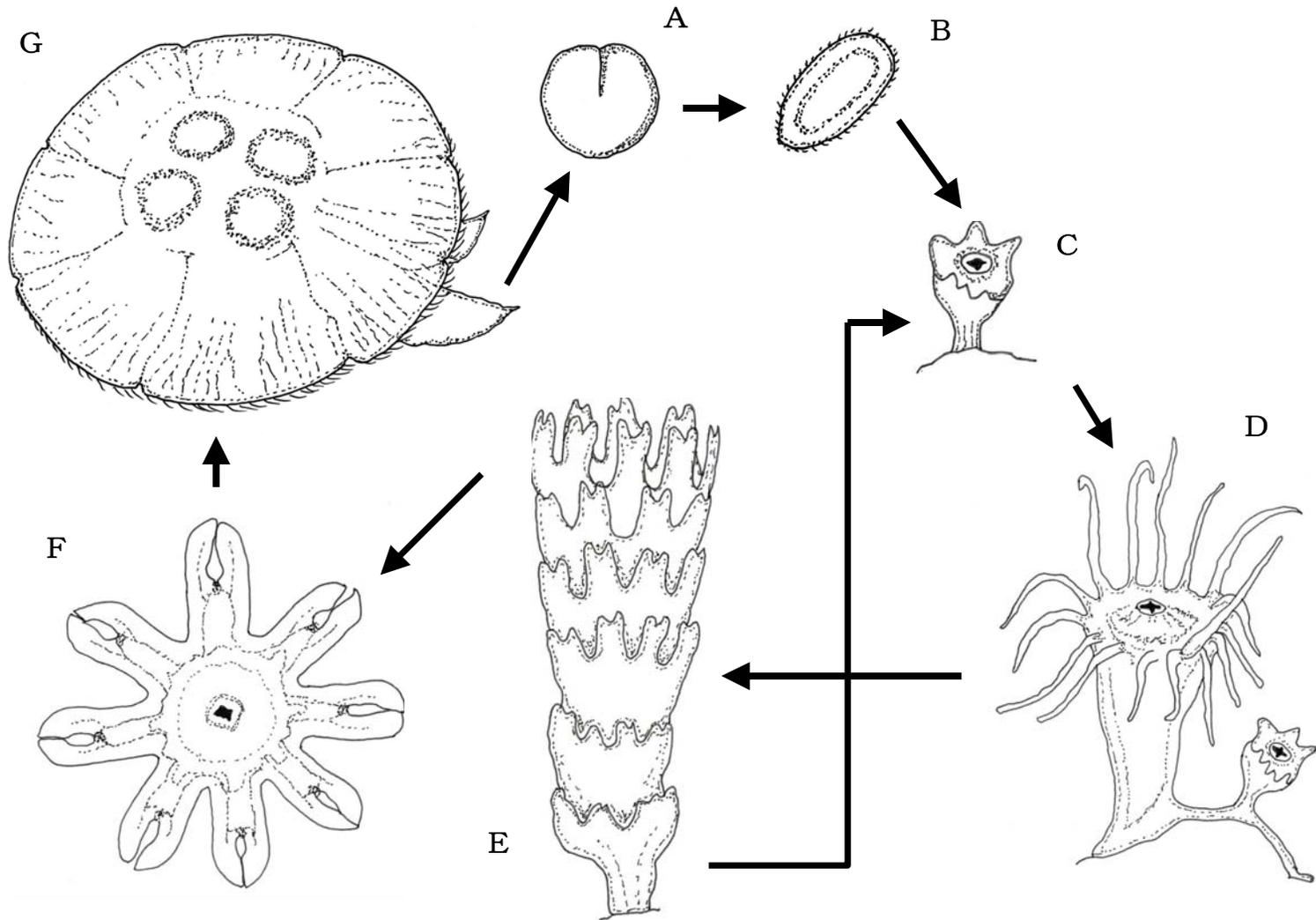


## Jelly Food Chain

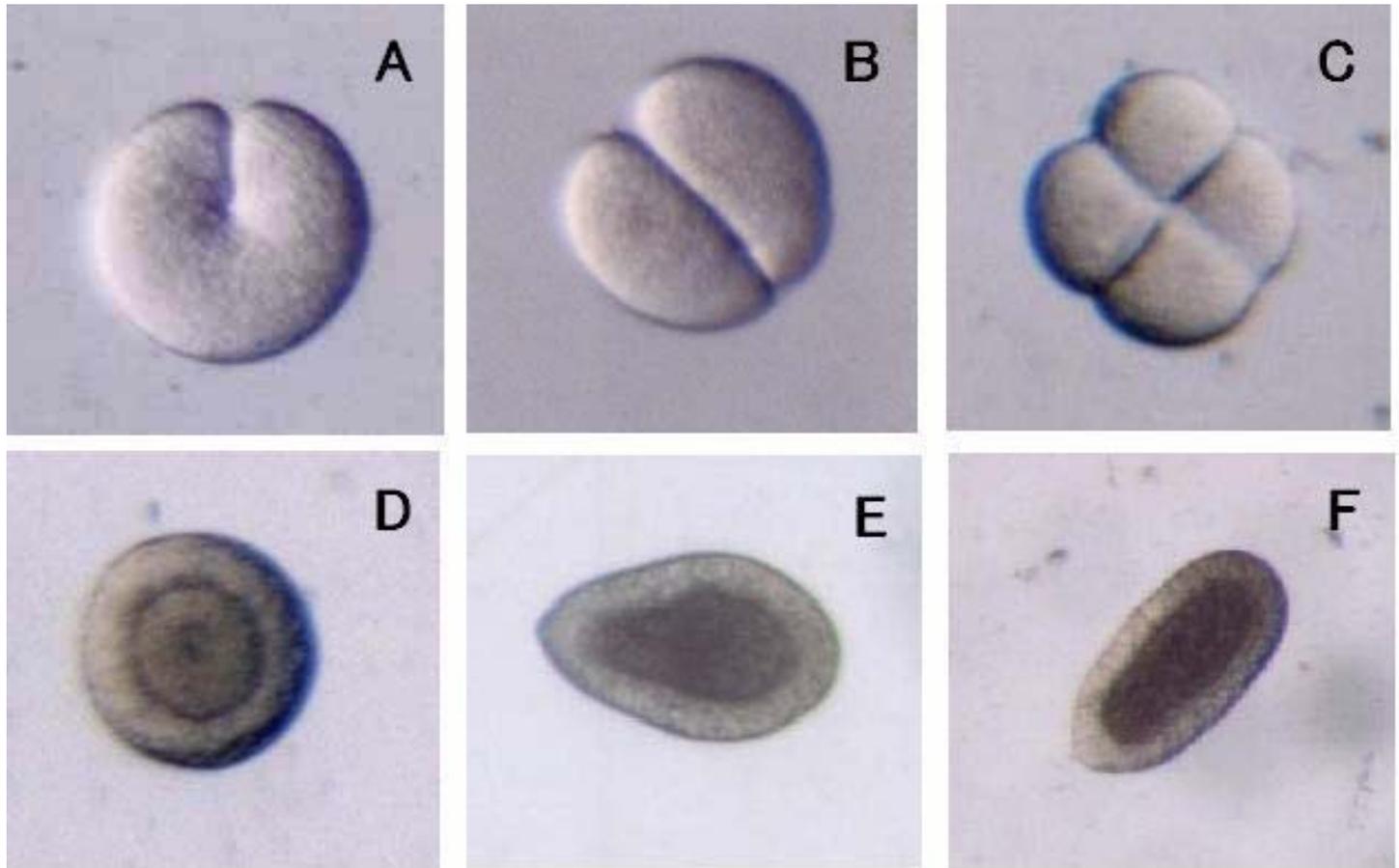
Microflagellate → Small zooplankton → Jellyfish

(Observed mainly in eutrophicated coastal systems)

Relationship between changes in coastal environment and mass occurrence of jellyfish in eutrophicated bay will be explained based on knowledge about phytoplankton responses to nutrients and food size spectra of zooplankton



**The life cycle**, including the annual sexual medusa stage and the perennial asexual polyp or scyphistoma stage: (A) first cleavage of zygote, (B) planula, (C) polyp, (D) asexual budding of daughter polyp, (E) strobila, (F) ephyra, and (G) medusa



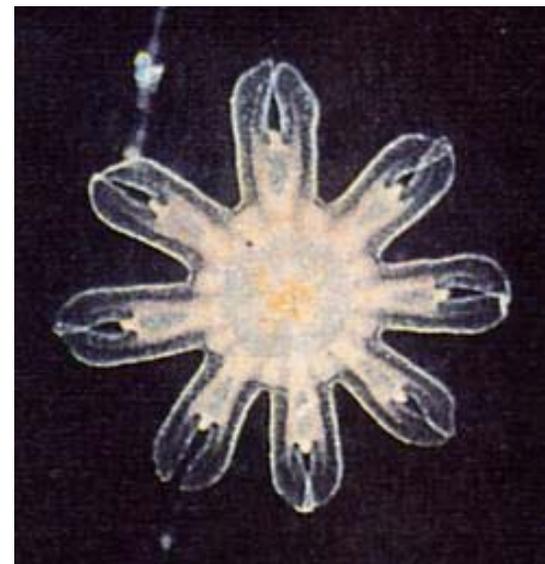
(A) first cleavage of **zygote** - (F) **planula**



**Polyp** with asexual budding of daughter polyp



**Strobila**

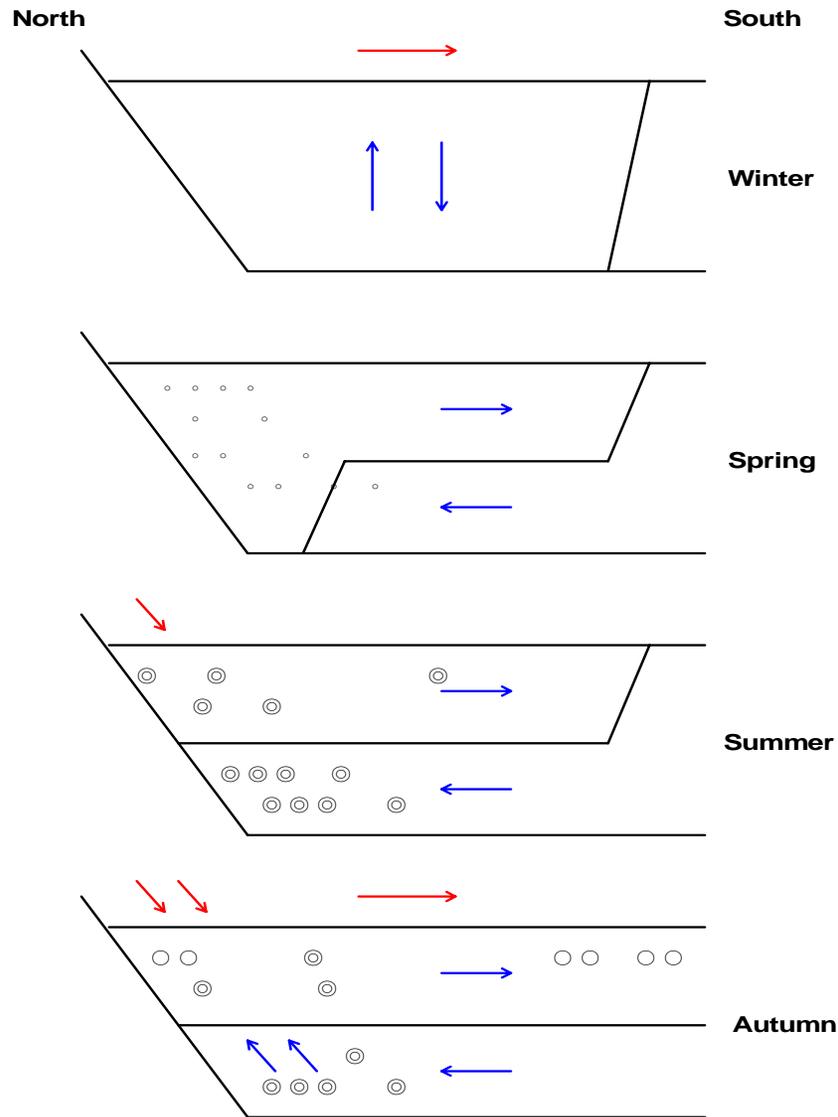


**Ephyra**



**Medusa**

# Seasonal population changes of *A. aurita* in Tokyo Bay



- **Ephyra**
- ◎ **Medusa**
- **Dead medusa**



Tokyo Bay

# Changes of the salinity and nutrient concentration in Tokyo Bay;

1950s ~ 2000s

Salinity decreasing (30.6 → 29.5 PSU)

Phosphate having a peak in 1970s (2 → 1  $\mu\text{g-at l}^{-1}$ )

Ammonium increasing (5 → 20  $\mu\text{g-at l}^{-1}$ )

Nitrite increasing (1 → 4  $\mu\text{g-at l}^{-1}$ )

Nitrate increasing (5 → 25  $\mu\text{g-at l}^{-1}$ )

Silicate decreasing (60 → 20  $\mu\text{g-at l}^{-1}$ )

## Increasing **N** and Decreasing **Si**

- Increasing river water and drainage → Eutrophication (**N** increasing)
- Dam construction, river improvement, coastal reclamation (disappearance of tideland) → **Si** decreasing
- Blooming of phytoplankton (Diatom Akashio event) → **Si** decreasing



# Low levels Si / N change phytoplankton composition; **Diatom** → **Microflagellate**

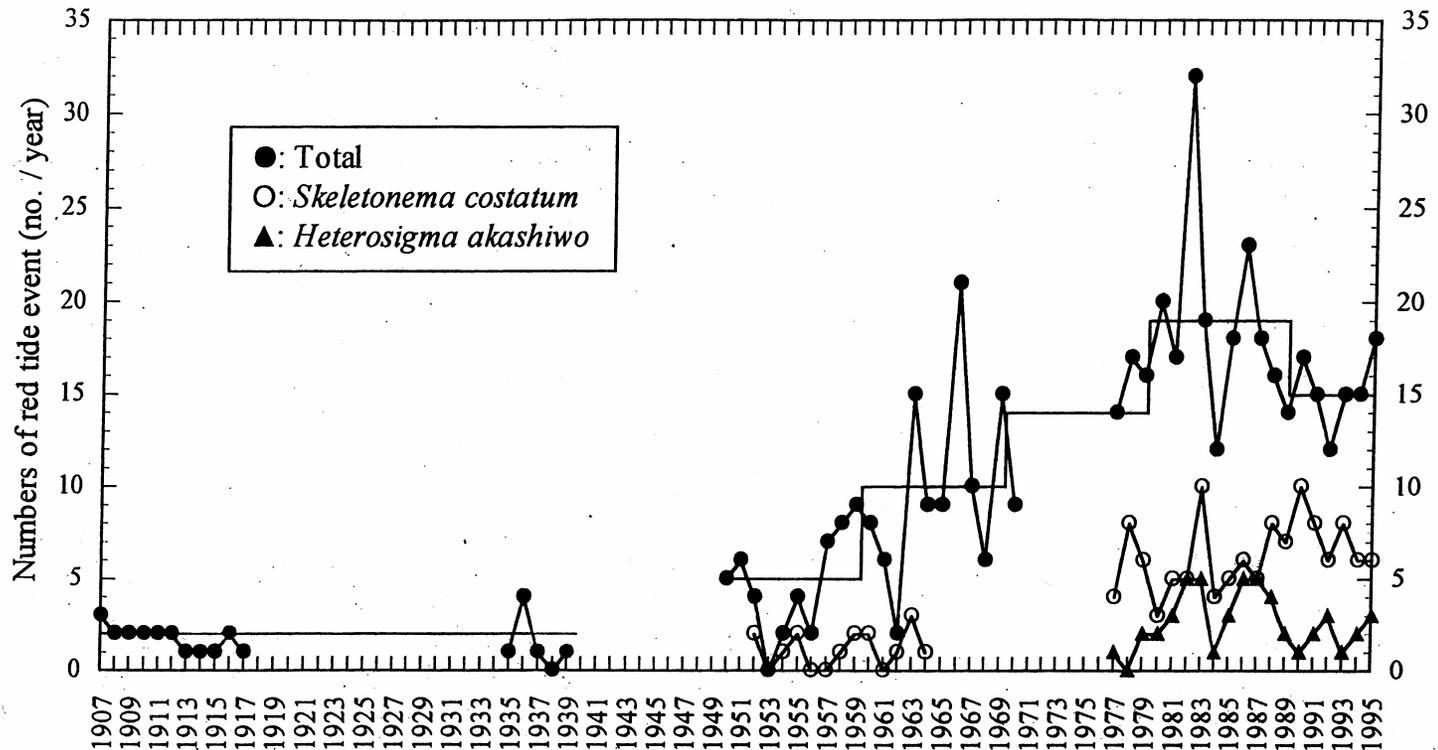
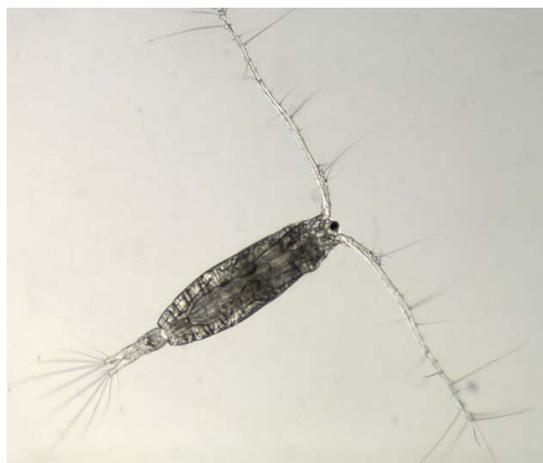


Fig. Changes in annual occurrence of total red tide events and the events by two main causative species (No./year) in Tokyo Bay from 1907 to 1995. Steplike line indicates the average number of red tide events in each period. Data from Asakura (1915), Nagata (1917), and Matsue (1935), Sugawara and Sato (1966a), Murata (1970), Tokyo-to Kankyo-hozen-kyoku Suishitsu-hozen-bu (1990-1997).

Nomura (1998)

# Zooplankton community; Meso- → Small-

Abundance ( $\times 10^3$ inds $m^{-3}$ )	1948	1971-72	1971-75	1979	1981-82	1983	2003
<i>Oithona</i>	515	470	600	1177	2045		1664
<i>Acartia</i>	28-38				23	13.2	1.9
	Yamaji (1955)	Marumo & Murano (1974)	Nishida (1985)	Nagasawa & Murano (1984)	Anakubo & Murano (1991)	Tsuda & Nemoto (1988)	Ishii unpubl.



Copepod (*Acartia omorii*)

Body length; 1 mm

Diatom feeder



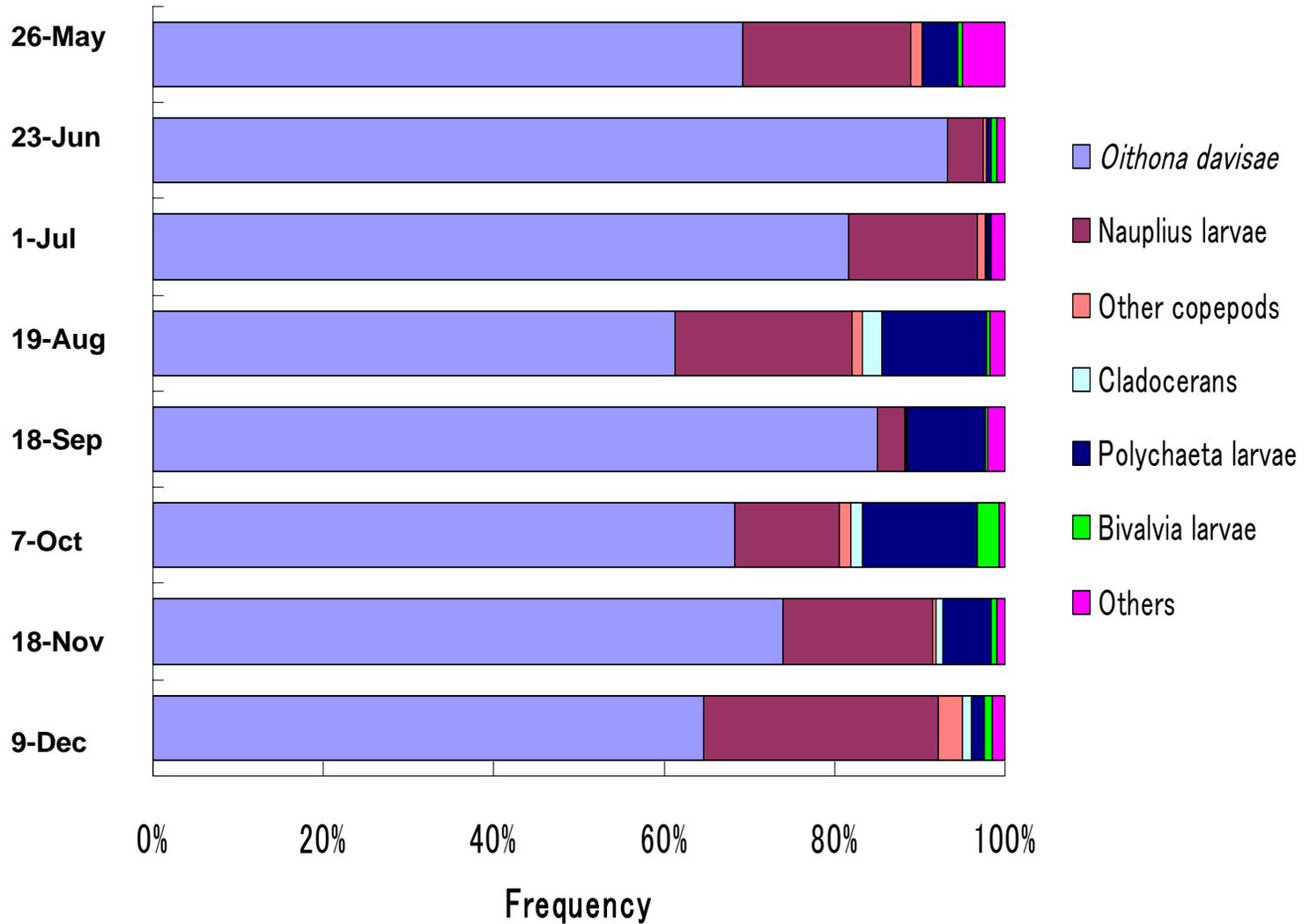
Small copepod

(*Oithona davisae*)

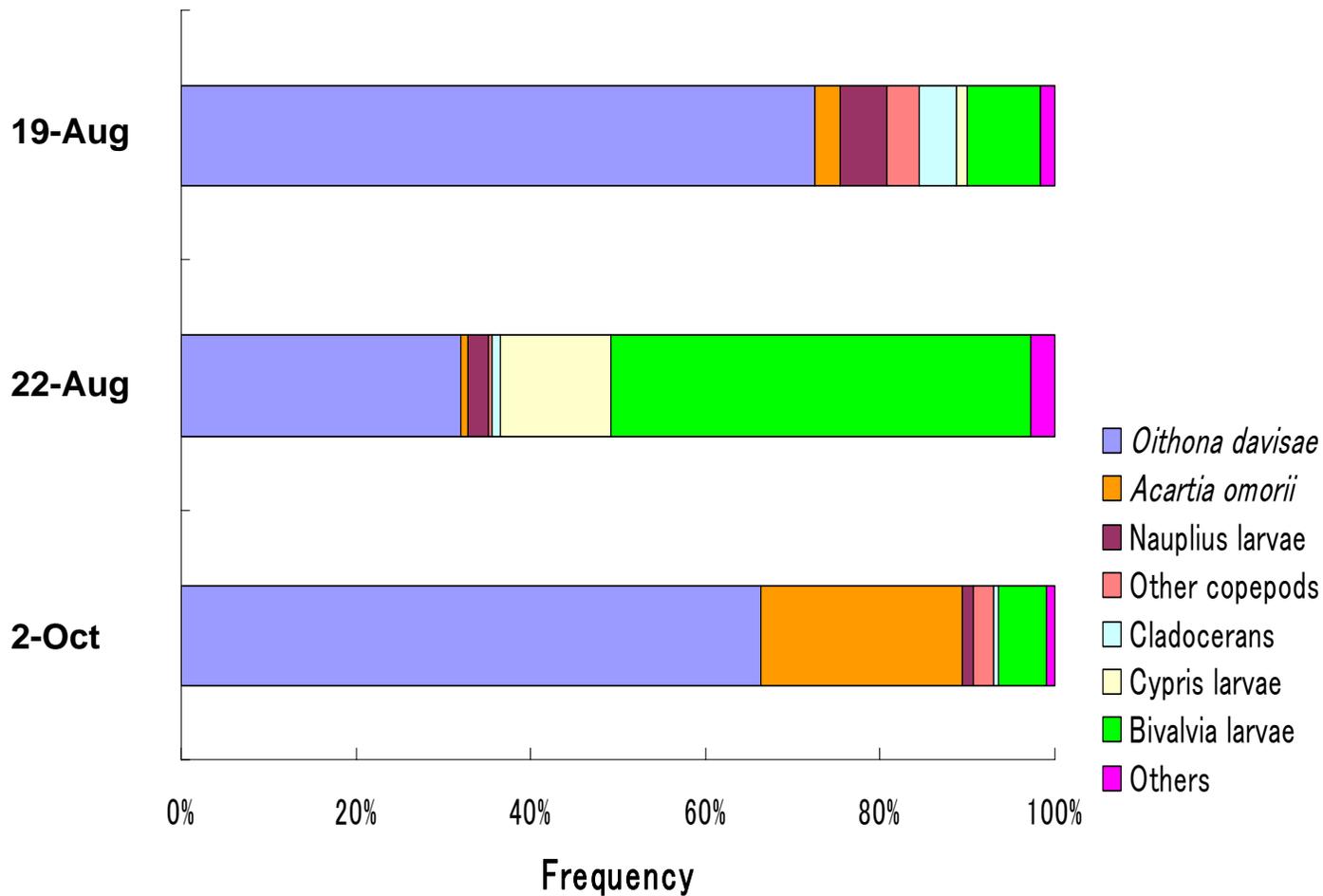
Body length; 0.3 mm

Microflagellate feeder

# Zooplankton composition in the seawater



# Prey composition in the stomach of medusae





# Jellyfish

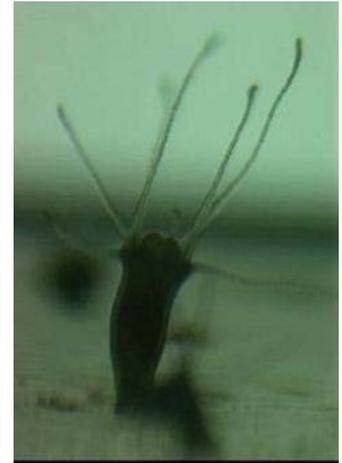
- Increase of the small prey abundance is suitable for jellyfish which are **opportunistic tactile predator**

# Fish

- Decrease of the large prey abundance and prey visibility is critical for planktivorous fish which are **visual predator**

Increase in the abundance of small copepod, *O. davisae*, have contributed to increases in jellyfish biomass relative to fish biomass

**Polyp stage** is characterized by asexual reproduction (budding, strobilation)

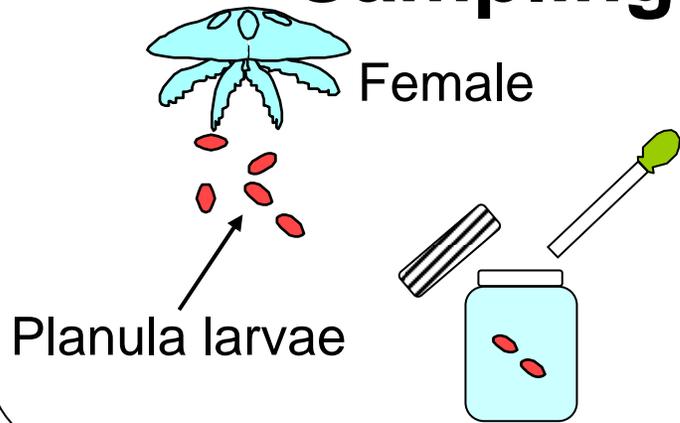


In coastal environment, settling substrate for polyp is increasing;

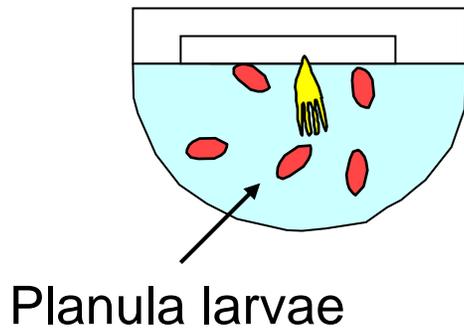
- Reclamation
- Quay
- Bottom of a ship



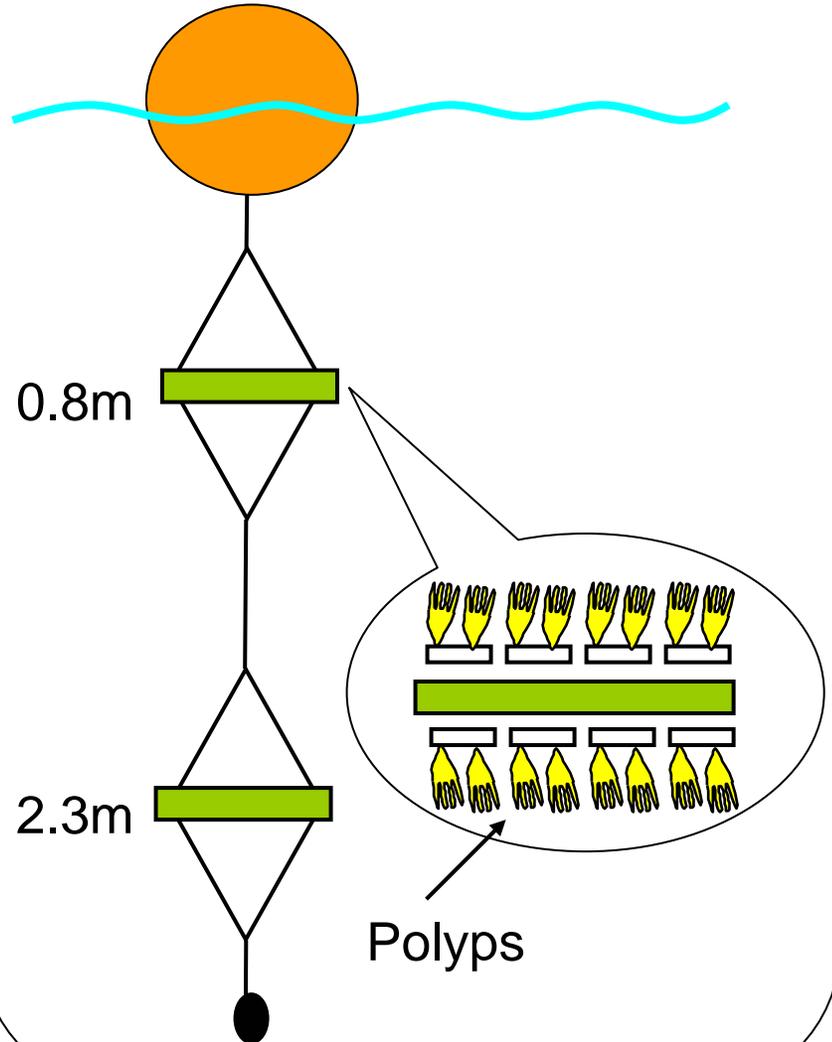
# Sampling

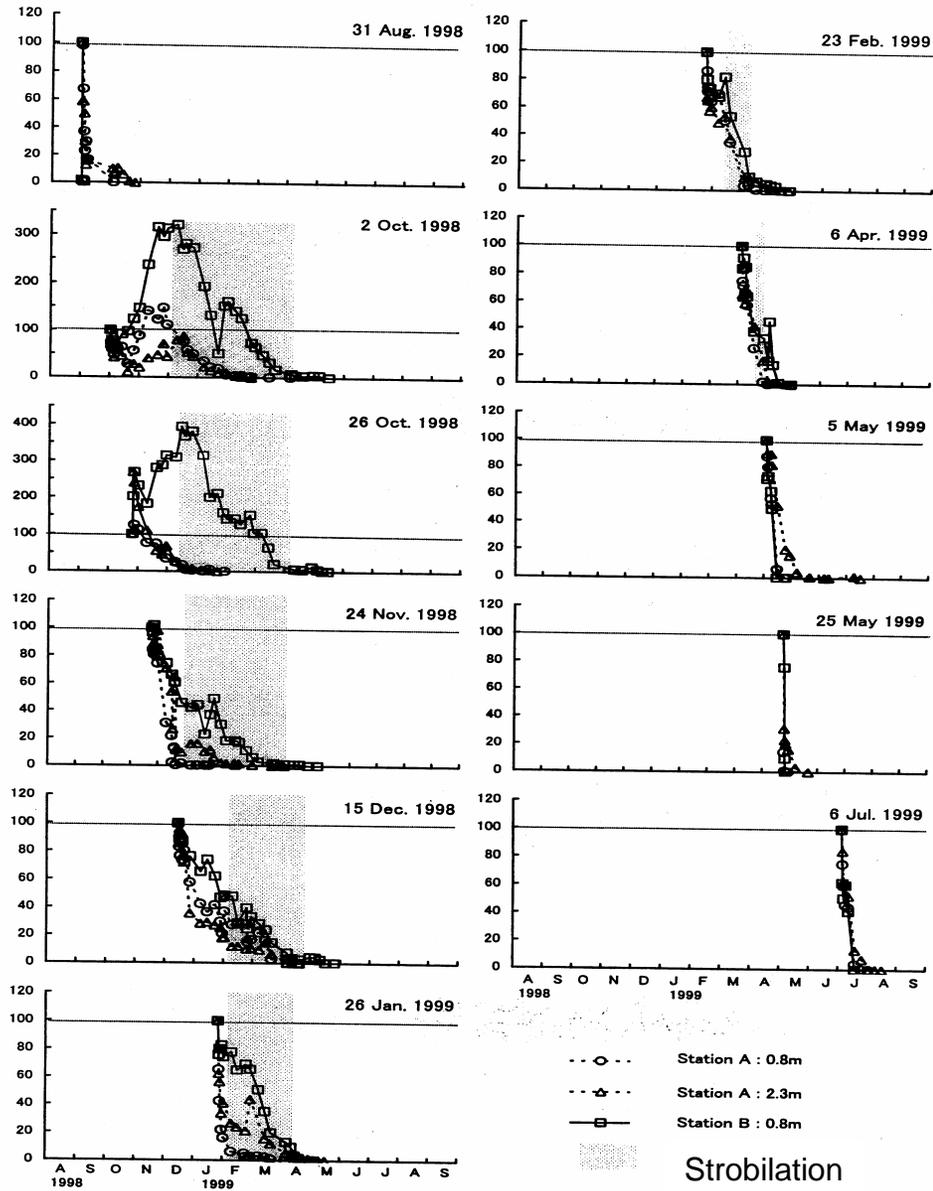


# Incubation



# Experiment





**Survivorship of polyps on the settling plates (%)**

Most of the settled substrate in the innermost part of Tokyo Bay is occupied by the other benthic organisms such as *Mitilus galloprovincialis*.

This observation means that *A. aurita* polyps are exposed to keen competition for space with other organisms, especially during spring and summer.

If the recruitment of planula larvae is restricted to summer, the consequent ephyra production could be low in the following spring.

The presence of ripe medusae with planulae, even in autumn and winter, would contribute to increasing settlement and survival of polyps.



Dense aggregation of  
*M. galloprovincialis*.  
No polyps are  
observed.

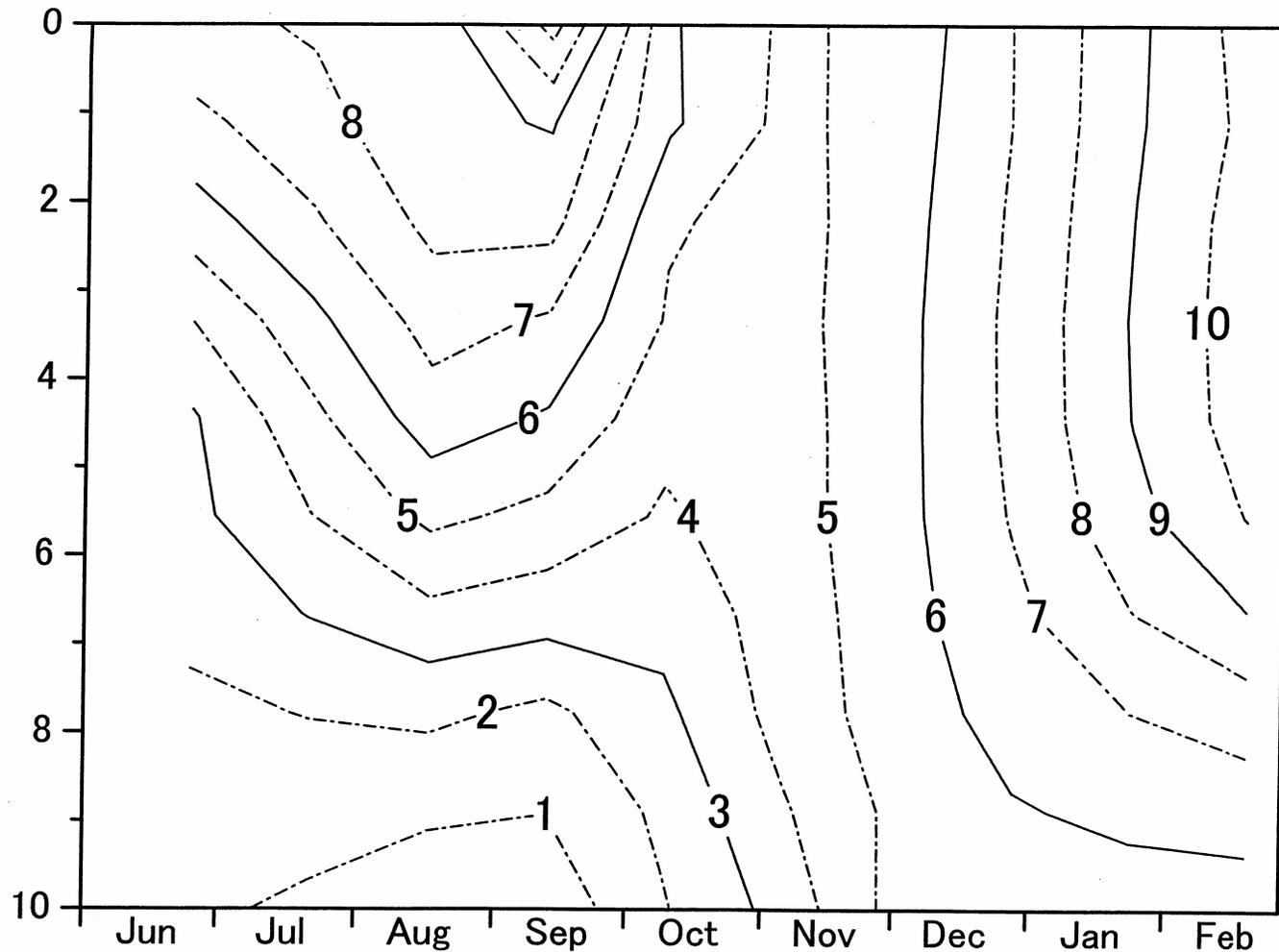
**We found the polyp aggregations in the **dysoxic bottom water** during summer (Peak season of planula release).**

This layer is characterized by low recruitment and growth of other benthic organisms such as *M. galloprovincialis*, resulting an abundant settlement and high survival during the polyp stage.



Dense  
aggregation  
of *A. aurita*  
polyps.

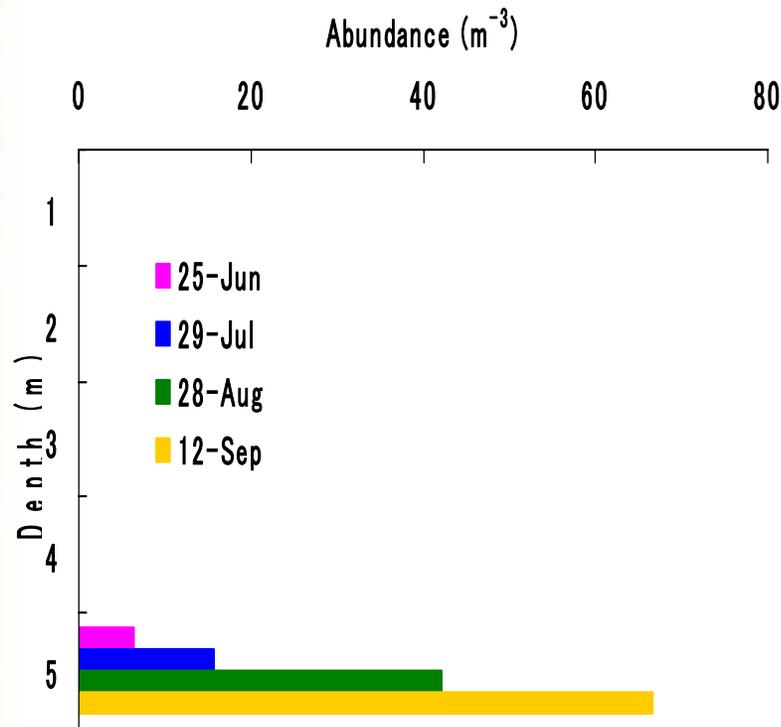
Depth (m)



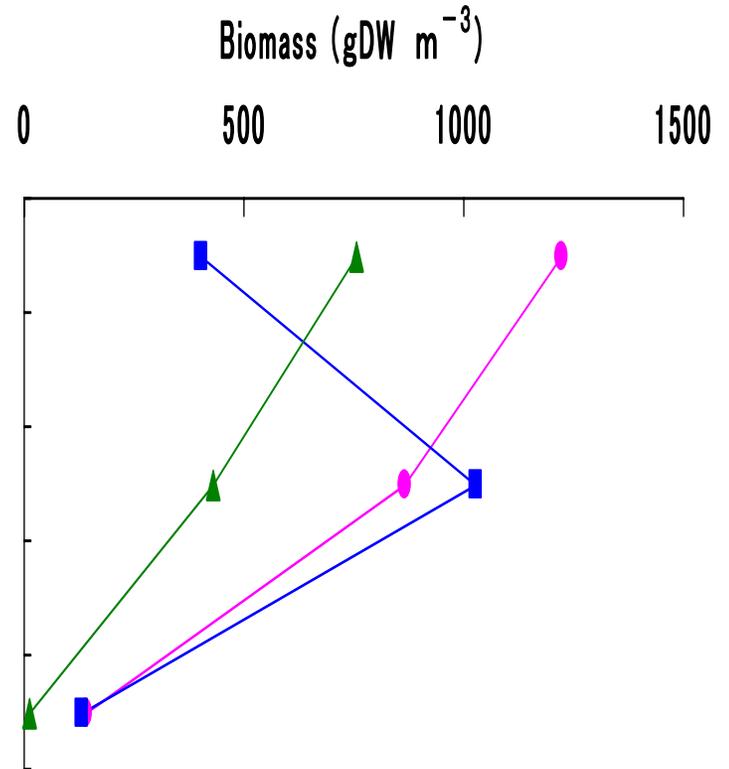
Seasonal changes in the concentration of D.O. (ml O<sub>2</sub> l<sup>-1</sup>) in the innermost part of Tokyo Bay.



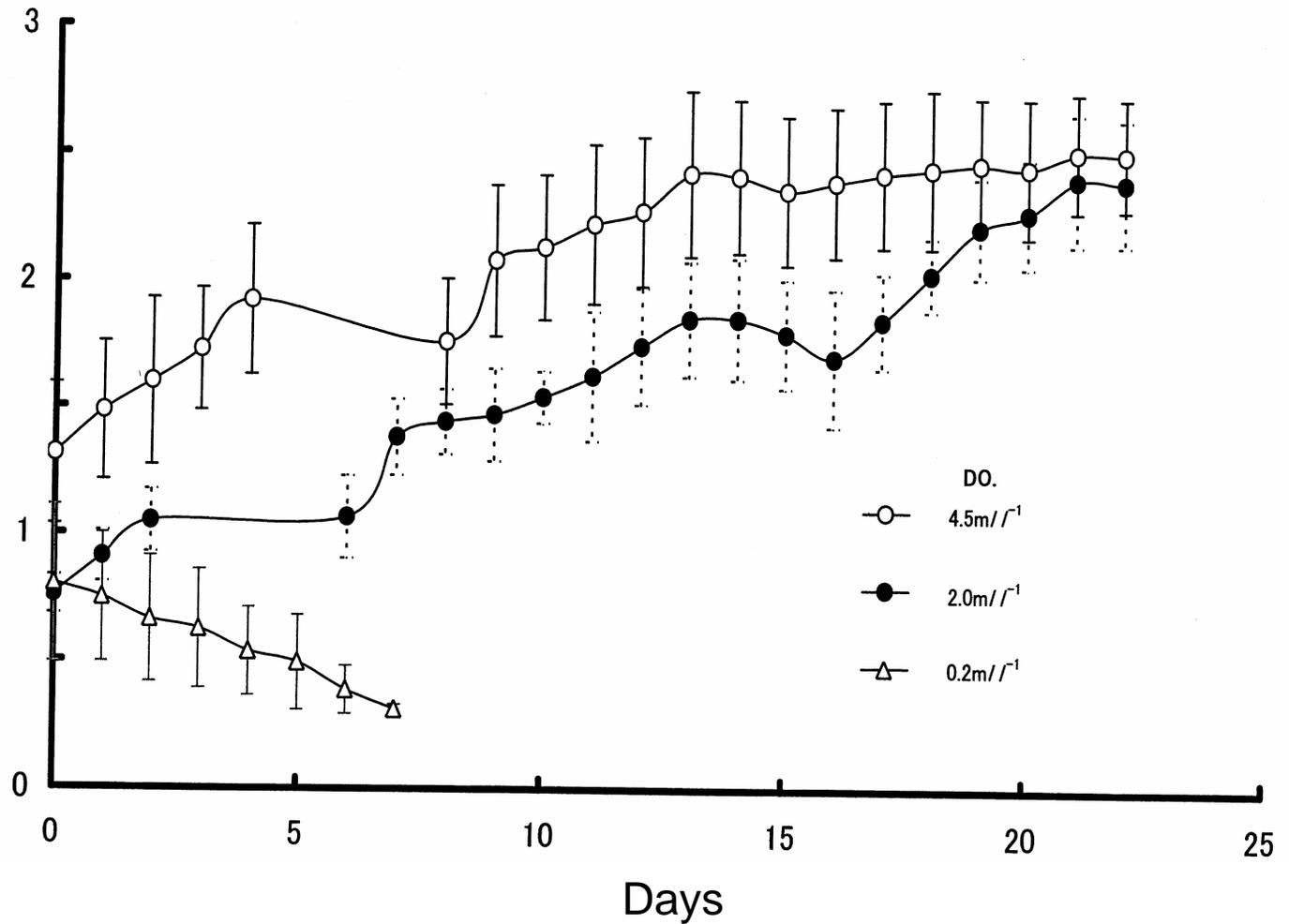
## Polyps of *A. aurita*



## *M. galloprovincialis*

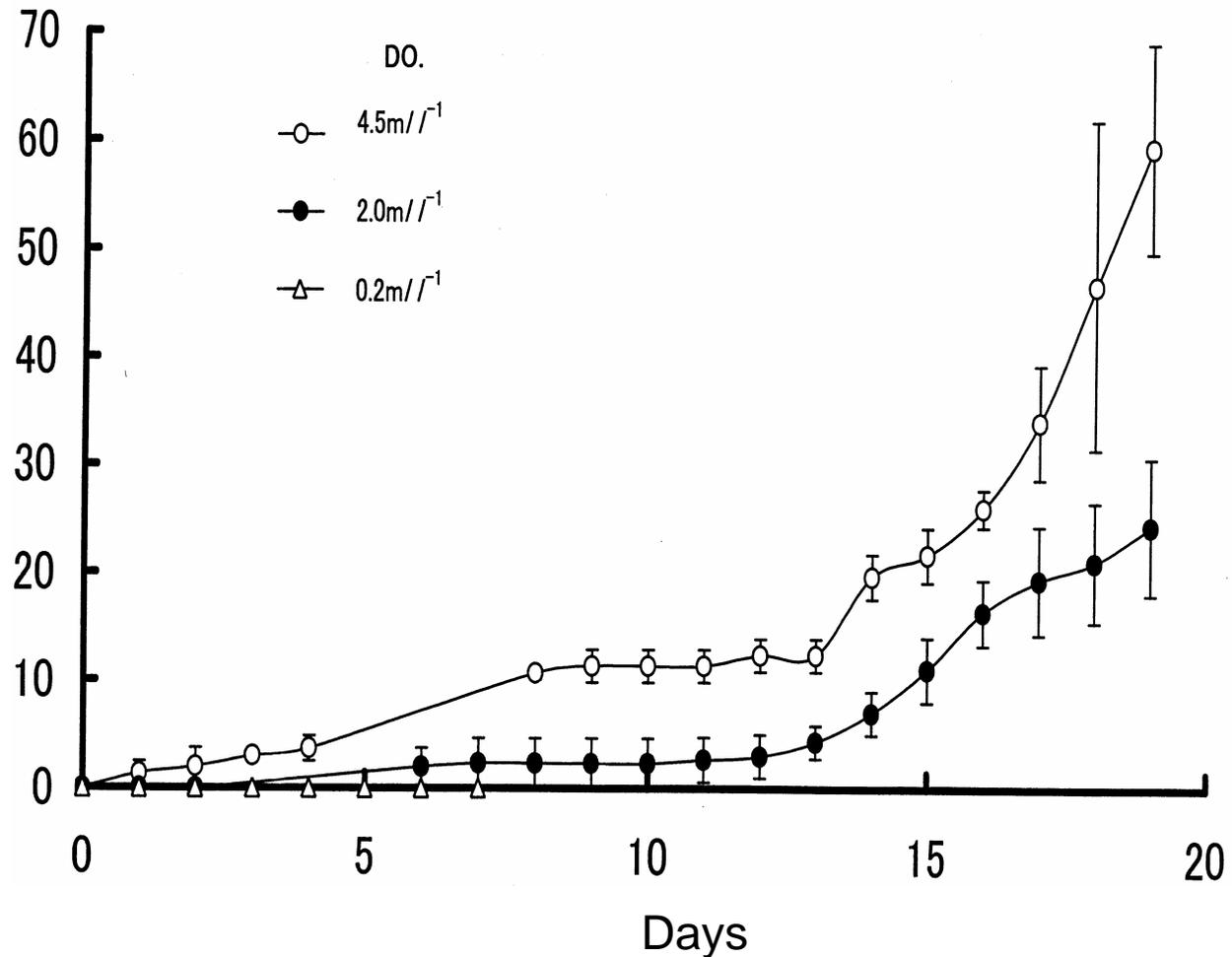


Polyp size (mm)



**Growth of polyps in the different D.O. concentration waters**

## Number of individuals



**Increase of daughter polyps by asexual budding in the different D.O. concentration waters**



# Polyp stage

When the D.O. concentration increased, strobilation and ephyra release are also observed even in the polyps cultured in hypoxic waters.

We still need to understand the relationship between **the abundance of polyps** with various jellyfish in eutrophic coastal waters and **their adaptation to dysoxic bottom-layer waters resulting from eutrophication.**



## **Guideline to prevent the mass occurrence of jellyfish**

### **1. Recover from hyper eutrophicated bay**

**Decreasing the nutrient (N and P) concentration  
→ Disappearance of red tide event and bottom dysoxic waters**

### **2. Balance of Si / N ratio in waters**

**Reconsideration of dam construction, river improvement, and coastal reclamation  
→ Increasing the size in phytoplankton (diatoms) and zooplankton (mesozooplankton) community  
→ Decreasing the polyp settling substrate**

**Since we cannot control the jellyfish biomass by top down method (no dominant predator), bottom up control will be effective.**

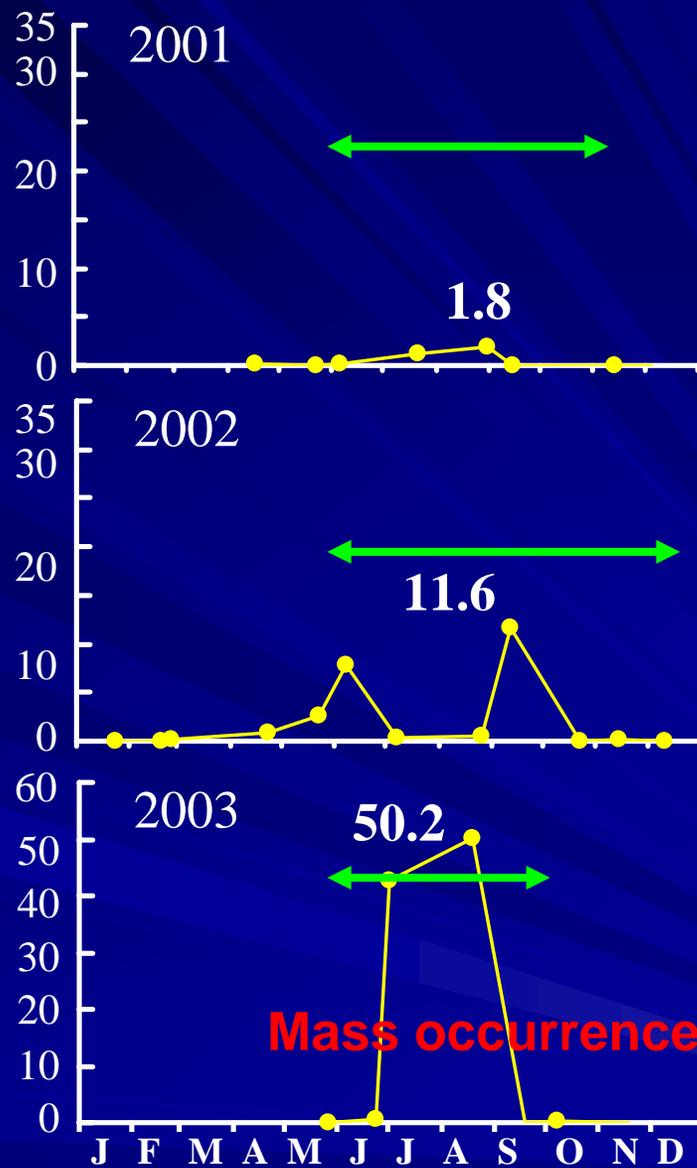
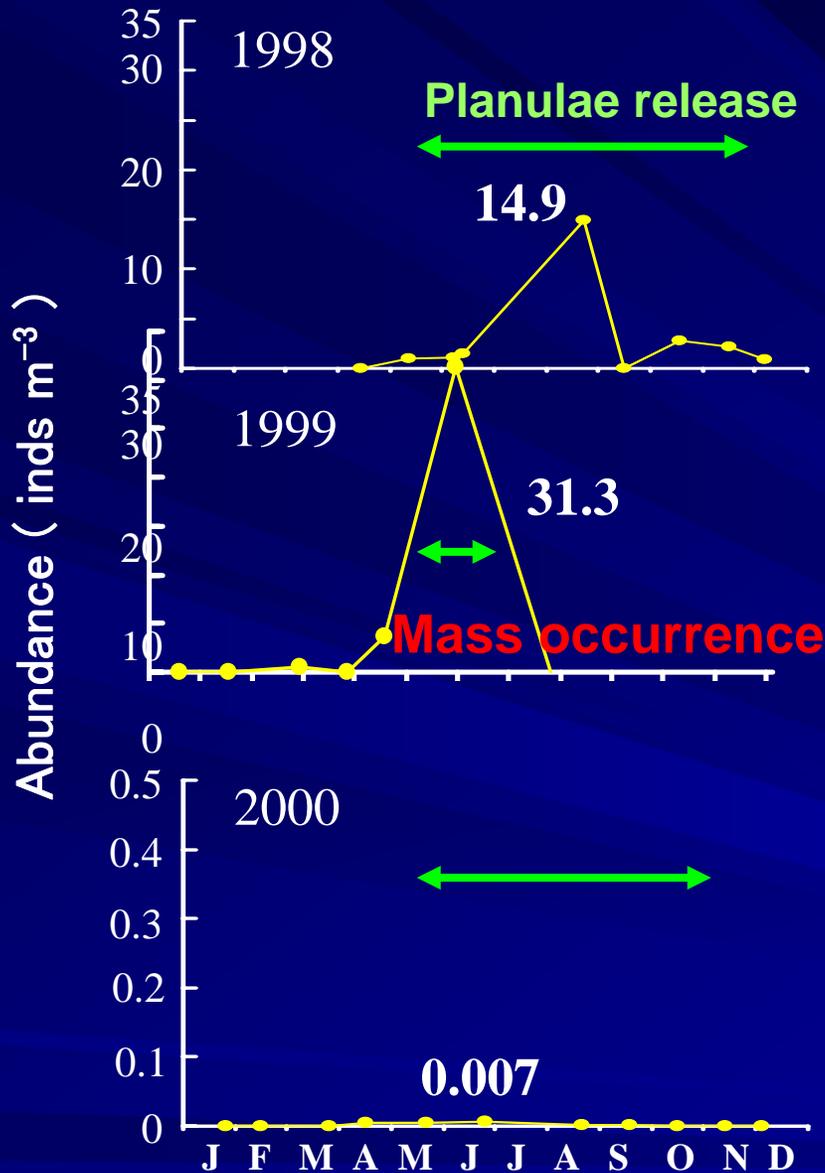
# Transition of Ecosystem

Muscle Food Chain (Fish) → Jelly Food Chain

Mass occurrence of gelatinous plankton including jellyfish with environmental changes is a symbolic event decreasing the biodiversity in marine ecosystem

## **Study:**

- **Construction of the simulation model including various parameters in relation to environmental changes; climate, river water, economic activity in the city, development of coastal region, eutrophication, and jellyfish biomass**
- **Prediction of the mass occurrence of jellyfish and control of the transition to jelly-ecosystem**



**Abundance of jellyfish, *A. aurita* in Tokyo Bay (1998~2003)**